Galectin 9 and 10SV Polypeptides

~~Galectin 8, 9, 10 and 10SV~~

~~This application claims the benefit of the filing date of provisional application 60/028,093 filed on October 9, 1996, which is herein incorporated by reference~~.

Background of the Invention

Field of the Invention

The present invention relates to novel galectins. More specifically, isolated nucleic acid molecules are provided encoding human galectin 8, 9, 10, or 10SV. Galectin 8, 9, 10 and 10SV polypeptides are also provided, as are vectors, host cells and recombinant methods for producing the same. The invention further relates to screening methods for identifying agonists and antagonists of galectin 8, 9, 10, or 10SV activity. Also provided are diagnostic methods for detecting cell growth disorders and therapeutic methods for cell growth disorders, including autoimmune diseases, cancer, and inflammatory diseases.

Related Art

Lectins are proteins that bind to specific carbohydrate structures and can thus recognize particular glycoconjugates. Barondes *et al., J. Biol. Chem. 269(33):*20807-20810 (1994). Galectins are members of a family of β-galactoside-binding lectins with related amino acid sequences (For review see, Barondes *et al., Cell 76:*597-598 (1994); Barondes *et al., J. Biol. Chem. 269(33):*20807-20810 (August 1994)). Galectin 1 (aka. L-14-1, L-14, RL-14.5, galaptin, MGBP, GBP, BHL, CHA, HBP, HPL, HLBP 14, rIML-1) is a homodimer with a subunit molecular mass of 14,500 which is abundant in smooth and skeletal muscle, and is present in many other cell types (Couraud *et al., J. Biol. Chem. 264:*1310-1316 (1989)). Galectin 2 was originally found in

hepatoma and is a homodimer with a subunit molecular weight of 14,650 (Gitt *et al., J. Biol. Chem. 267:*10601-10606 (1992)). Galectin 3 (aka. Mac-2, EPB, CBP-35, CBP-30, and L-29) is abundant in activated macrophages and epithelial cells and is a monomer with an apparent molecular mass between 26,320 and 30,300 (Cherayil *et al., Proc. Natl. Acad. Sci. USA 87:* 7324-7326 (1990)). Galectin 4 has a molecular mass of 36,300 and contains two carbohydrate-binding domains within a single polypeptide chain (Oda *et al., J. Biol. Chem. 268:*5929-5939 (1993)). Galectins 5 and 6 are mentioned in Barondes *et al., Cell 76:*597-598 (1994). Human galectin 7 has a molecular mass of 15,073 and is found mainly in stratified squamous epithelium (Madsen *et al., J. Biol. Chem. 270(11):*5823-5829 (1995)).

Animal lectins, in general, often function in modulating cell-cell and cell-matrix interactions. Galectin 1 has been shown to either promote or inhibit cell adhesion depending upon the cell type in which it is present. Galectin 1 inhibits cell-matrix interactions in skeletal muscle (Cooper *et al., J. Cell Biol. 115:*1437-1448 (1991)). In other cell types, galectin 1 promotes cell-matrix adhesion possibly by cross-linking cell surface and substrate glycoconjugates (Zhou *et al., Arch. Biochem. Biophys. 300:*6-17 (1993); Skrincosky *et al., Cancer Res. 53:*2667-2675 (1993)).

Galectin 1 also participates in regulating cell proliferation (Wells *et al., Cell 64:*91-97 (1991)) and some immune functions (Offner *et al., J. Neuroimmunol. 28:*177-184 (1990)). Galectin 1 has been shown to regulate the immune response by mediating apoptosis of T cells (Perillo *et al., Nature 378:* 736-739 (1995)).

Galectin 3 promotes the growth of cells cultured under restrictive culture conditions (Yang *et al., Proc. Natl. Acad. Sci. USA 93:*6737-6742 (June 1996)). Galectin 3 expression in cells confers resistance to apoptosis which indicates that Galectin 3 could be a cell death suppressor which interferes in a common pathway of apoptosis. *Id.*

Accordingly, there is a need in the art for the identification of novel galectins which can serve as useful tools in the development of therapeutics and diagnostics for regulating immune response.

Summary of the Invention

The present invention provides isolated nucleic acid molecules comprising a polynucleotide encoding the galectin 8, 9, 10, or 10SV polypeptide having the amino acid sequence is shown in FIGs. 1, 2A-2B, 3A-3B, and 4A-4B, respectively (SEQ ID NOs:2, 4, 6, and 8, respectively) or the amino acid sequence encoded by the cDNA clones deposited ~~in bacterial hosts~~ as ATCC Deposit Numbers 97732, 97733 and 97734 on September 24, 1996.

The present invention also relates to recombinant vectors, which include the isolated nucleic acid molecules of the present invention, and to host cells containing the recombinant vectors, as well as to methods of making such vectors and host cells and for using them for production of galectin 8, 9, 10, or 10SV polypeptides or peptides by recombinant techniques.

The invention further provides an isolated galectin 8, 9, 10, or 10SV polypeptide having an amino acid sequence encoded by a polynucleotide described herein.

The present invention also provides a screening method for identifying compounds capable of enhancing or inhibiting a cellular response induced by galectin 8, 9, 10, or 10SV, which involves contacting cells which express galectin 8, 9, 10, or 10SV with the candidate compound, assaying a cellular response, and comparing the cellular response to a standard cellular response, the standard being assayed when contact is made in absence of the candidate compound; whereby, an increased cellular response over the standard indicates that the compound is an agonist and a decreased cellular response over the standard indicates that the compound is an antagonist.

In another aspect, a screening assay for agonists and antagonists is provided which involves determining the effect a candidate compound has on galectin 8, 9, 10, or 10SV binding to the β-galactosidase sugar. In particular, the method involves contacting the β-galactosidase sugar with a galectin 8, 9, 10, or 10SV polypeptide and a candidate compound and determining whether galectin 8, 9, 10, or 10SV binding to β-galactosidase sugar is increased or decreased due to the presence of the candidate compound.

The invention provides a diagnostic method useful during diagnosis disorder.

An additional aspect of the invention is related to a method for treating an individual in need of an increased level of galectin 8, 9, 10, or 10SV activity in the body comprising administering to such an individual a composition comprising a therapeutically effective amount of an isolated galectin 8, 9, 10, or 10SV polypeptide of the invention or an agonist thereof.

A still further aspect of the invention is related to a method for treating an individual in need of a decreased level of galectin 8, 9, 10, or 10SV activity in the body comprising, administering to such an individual a composition comprising a therapeutically effective amount of a galectin 8, 9, 10, or 10SV antagonist.

Brief Description of the Figures

FIG. 1 shows the nucleotide (SEQ ID NO:1) and deduced amino acid (SEQ ID NO:2) sequences of galectin 8. The protein has a deduced molecular weight of about 36 kDa.

FIG. 2A-2B shows the nucleotide (SEQ ID NO:3) and deduced amino acid (SEQ ID NO:4) sequences of galectin 9. The protein has a deduced molecular weight of about 34.7 kDa.

FIG. 3A-3B shows the nucleotide (SEQ ID NO:5) and deduced amino acid (SEQ ID NO:6) sequences of full length galectin 10. The protein has a deduced molecular weight of about 35.7 kDa.

FIG. 4A-4B shows the nucleotide (SEQ ID NO:7) and deduced amino acid (SEQ ID NO:8) sequences of a galectin 10 splice variant (galectin 10SV). The protein has a deduced molecular weight of about 22.4 kDa.

FIG. 5A-~~5B~~ 5E shows the regions of similarity between the amino acid sequences of the galectin 8, 9, and 10 proteins and human galectin 2 (SEQ ID NO:9), human galectin 3 (SEQ ID NO:10), rat galectin 4 (SEQ ID NO:11), rat galectin 5 (SEQ ID NO:12), human galectin 7 (SEQ ID NO:13), rat galectin 3 (SEQ ID NO:14), rat galectin 8 (SEQ ID NO:15), and human galectin 1 (SEQ ID NO:16).

FIG. 6 shows the regions of similarity between the amino acid sequences of the galectin 10SV protein and the rat RL30 protein (SEQ ID NO:17).

FIG. 7 shows a homology comparison between the galectin 10 and galectin 10SV proteins.

FIGs. 8, 9, 10, and 11 show an analysis of the galectin 8, 9, 10, and 10SV amino acid sequence, respectively. Alpha, beta, turn and coil regions; hydrophilicity and hydrophobicity; amphipathic regions; flexible regions; antigenic index and surface probability are shown. In the "Antigenic Index - Jameson-Wolf" graph, amino acid residues 55-101, 137-162, 180-193, 216-266 in FIG. 1 (SEQ ID NO:2), 62-102, 226-259, 197-308 in FIG. 2A-2B (SEQ ID NO:4), 25-77, 84-105, 129-140, 156-183, 195-215, and 241-257 in FIG. 3A-3B (SEQ ID NO:6), and 25-77, 84-105, 129-140, and 156-183 in FIG. 4A-4B (SEQ ID NO:8) correspond to the shown highly antigenic regions of the galectin 8, 9, 10, or 10SV protein, respectively.

Detailed Description

The present invention provides isolated nucleic acid molecules comprising a polynucleotide encoding a galectin 8, 9, 10, or 10SV polypeptide having the amino acid sequence shown in FIGs. 1, 2A-2B, 3A-3B, and 4A-4B, respectively (SEQ ID NOs:2, 4, 6, and 8, respectively), which was determined by sequencing

a cloned cDNA. The galectin 8, 9, 10, and 10SV proteins of the present invention share sequence homology with other galectins and the rat RL30 protein (FIGs. 5A-5B and 6) (SEQ ID NOs:9-17). The nucleotide sequences shown in FIGs. 1, 2A-2B, and 4A-4B (SEQ ID NO:1, 3, and 7, respectively) were obtained by sequencing the HSIAL77, HTPBR22, and HETAS87 clones, which were deposited on September 24, 1996 at the American Type Culture Collection, 10801 University Blvd., Manassas, VA 20110-2209, USA ~~12301 Park Lawn Drive, Rockville, Maryland 20852~~, and given accession numbers 97732, 97733 and 97734, respectively. The deposited clones are contained in the pBluescript SK(-) plasmid (Stratagene, LaJolla, CA).

The nucleotide sequence shown in FIG. 3A-3B (SEQ ID NO:5), which encodes the full-length galectin 10 protein, was obtained by sequencing a clone cDNA obtained from a human endometrial tumor library.

Nucleic Acid Molecules

Unless otherwise indicated, all nucleotide sequences determined by sequencing a DNA molecule herein were determined using an automated DNA sequencer (such as the Model 373 from Applied Biosystems, Inc.), and all amino acid sequences of polypeptides encoded by DNA molecules determined herein were predicted by translation of a DNA sequence determined as above. Therefore, as is known in the art for any DNA sequence determined by this automated approach, any nucleotide sequence determined herein may contain some errors. Nucleotide sequences determined by automation are typically at least about 90% identical, more typically at least about 95% to at least about 99.9% identical to the actual nucleotide sequence of the sequenced DNA molecule. The actual sequence can be more precisely determined by other approaches including manual DNA sequencing methods well known in the art. As is also known in the art, a single insertion or deletion in a determined nucleotide sequence compared to the actual sequence will cause a frame shift in translation of the nucleotide sequence such that the predicted amino acid

sequence encoded by a determined nucleotide sequence will be completely different from the amino acid sequence actually encoded by the sequenced DNA molecule, beginning at the point of such an insertion or deletion.

Using the information provided herein, such as the nucleotide sequences in FIGs. 1, 2A-2B, 3A-3B, and 4A-4B a nucleic acid molecule of the present invention encoding a galectin 8, 9, 10, or 10SV, respectively, polypeptide may be obtained using standard cloning and screening procedures, such as those for cloning cDNAs using mRNA as starting material. Illustrative of the invention, the nucleic acid molecules described in FIGs. 1, 2A-2B, 3A-3B, and 4A-4B (SEQ ID NO:1, 3, 5, and 7, respectively) were discovered in cDNA libraries derived from human adult small intestine, human pancreatic tumor, human endometrial tumor and human endometrial tumor, respectively. These genes were also identified in cDNA libraries from the following tissues pancreas, colon, small intestine, brain, bone marrow, kidney, lung, spleen, and testes tissue. Galectin 8 (SEQ ID NO:1) appears to be mainly expressed in cells of the human colon and small intestine.

The determined nucleotide sequences of the galectin 8, 9, 10, and 10SV cDNAs of FIGs. 1, 2A-2B, 3A-3B, and 4A-4B, respectively (SEQ ID NOs:1, 3, 5, and 7) contain open reading frames encoding proteins of 323, 311, 317, and 200 amino acid residues, with an initiation codon at positions 52-54, 16-18, 118-120, and 118-120 of the nucleotide sequences in FIGs. 1, 2A-2B, 3A-3B, and 4A-4B, respectively (SEQ ID NOs:1, 3, 5, and 7), and a deduced molecular weight of about 36, 34.7, 35.7, and 22.4 kDa, respectively. The galectin 8, 9, 10 and 10SV proteins shown in FIGs. 1, 2A-2B, 3A-3B, and 4A-4B respectively (SEQ ID NOs:2, 4, 6, and 8) share homology with other galectins (See, *e.g.*, FIG. 5A-5B).

As one of ordinary skill would appreciate, due to the possibilities of sequencing errors discussed above, as well as the variability of processing sites for different known proteins, the predicted galectin 8 and 9 polypeptides encoded by the deposited cDNAs comprise about 323 and 311 amino acids, but may be

anywhere in the range of 300 - 333 amino acids. Similarly, the predicted galectin 10 polypeptide comprises about 317 amino acids, but may be anywhere in the range of 305 - 329 amino acids. Further, the predicted galectin 10SV polypeptide encoded by the deposited cDNA comprises about 200 amino acids, but may be anywhere in the range of 190 - 210 amino acids.

Galectin 10SV is believed to be a splice variant of galectin 10. As used herein the phrase "splice variant" refers to cDNA molecules produced from RNA molecules initially transcribed from the same genomic DNA sequence but which have undergone alternative RNA splicing. Alternative RNA splicing occurs when a primary RNA transcript undergoes splicing, generally for the removal of introns, which results in the production of more than one mRNA molecule each of which may encode different amino acid sequences. The term "splice variant" also refers to the proteins encoded by the above cDNA molecules.

As indicated, nucleic acid molecules of the present invention may be in the form of RNA, such as mRNA, or in the form of DNA, including, for instance, cDNA and genomic DNA obtained by cloning or produced synthetically. The DNA may be double-stranded or single-stranded. Single-stranded DNA or RNA may be the coding strand, also known as the sense strand, or it may be the non-coding strand, also referred to as the anti-sense strand.

By "isolated" nucleic acid molecule(s) is intended a nucleic acid molecule, DNA or RNA, which has been removed from its native environment For example, recombinant DNA molecules contained in a vector are considered isolated for the purposes of the present invention. Further examples of isolated DNA molecules include recombinant DNA molecules maintained in heterologous host cells or purified (partially or substantially) DNA molecules in solution. Isolated RNA molecules include *in vivo* or *in vitro* RNA transcripts of the DNA molecules of the present invention. Isolated nucleic acid molecules according to the present invention further include such molecules produced synthetically.

Isolated nucleic acid molecules of the present invention include DNA molecules comprising an open reading frame (ORF) shown in FIGs. 1, 2A-2B,

3A-3B, and 4A-4B, respectively (SEQ ID NOs:1, 3, 5, and 7); and DNA molecules which comprise a sequence substantially different from those described above but which, due to the degeneracy of the genetic code, still encode the galectin 8, 9, 10, or 10SV protein. Of course, the genetic code is well known in the art. Thus, it would be routine for one skilled in the art to generate such degenerate variants.

In addition, the invention provides nucleic acid molecules having nucleotide sequences related to extensive portions of SEQ ID NO:1 which have been determined from the following related cDNA clones: HSIAL77R (SEQ ID NO:18), HGBDK55R (SEQ ID NO:19), HCNAH29R (SEQ ID NO:20), HKCAA85R (SEQ ID NO:21), HCNAI55R (SEQ ID NO:22), HCNAI87R (SEQ ID NO:23), HCNAS74R (SEQ ID NO:24) and HCNAF43R (SEQ ID NO:25).

In addition, the invention provides nucleic acid molecules having nucleotide sequences related to extensive portions of SEQ ID NO:3 which have been determined from the following related cDNA clones: HMSCP11R (SEQ ID NO:26), HMSEU32R (SEQ ID NO:27), HTPAO71R (SEQ ID NO:28), HJAAV54R (SEQ ID NO:29), HMSEU43R (SEQ ID NO:30), HILBP03R (SEQ ID NO:31), HTPCG81R (SEQ ID NO:32), HTBAA21R (SEQ ID NO:33), and HFXBU26R (SEQ ID NO:34).

In addition, the invention provides nucleic acid molecules having nucleotide sequences related to extensive portions of SEQ ID NO:5 which have been determined from the following related cDNA clones: HTNBX92R (SEQ ID NO:35), HLTAZ64RB (SEQ ID NO:36), HJBAI38R (SEQ ID NO:37), HETAS87R (SEQ ID NO:38), and HETAR45R (SEQ ID NO:39).

In addition, the invention provides nucleic acid molecules having nucleotide sequences related to extensive portions of SEQ ID NO:7 which have been determined from the following related cDNA clones: HTNBX92R (SEQ ID NO:35), HLTAZ64RB (SEQ ID NO:36), HBNAF37R (SEQ ID NO:40), and HETAS87R (SEQ ID NO:38).

In another aspect, the invention provides isolated nucleic acid molecules encoding the galectin 8, 9, 10 or 10SV polypeptide having an amino acid sequence encoded by the cDNA clone contained in the plasmid deposited as ATCC Deposit Nos. 97732, 97733 and 97734, respectively, on September 24, 1996. In a further embodiment, nucleic acid molecules are provided encoding the full-length galectin 8, 9, 10, or 10SV polypeptide lacking the N-terminal methionine. The invention further provides an isolated nucleic acid molecule having the nucleotide sequence shown in FIGs. 1, 2A-2B, 3A-3B, or 4A-4B (SEQ ID NOs:1, 3, 5, or 7) or the nucleotide sequence of the galectin 8, 9, or 10SV cDNA contained in the above-described deposited clones, or a nucleic acid molecule having a sequence complementary to one of the above sequences. Such isolated molecules, particularly DNA molecules, are useful as probes for gene mapping, by *in situ* hybridization with chromosomes, and for detecting expression of the galectin 8, 9, 10, or 10SV gene in human tissue, for instance, by Northern blot analysis.

The present invention is further directed to fragments of the isolated nucleic acid molecules described herein. By a fragment of an isolated nucleic acid molecule having the nucleotide sequence of the deposited cDNA or the nucleotide sequence shown in FIGs. 1, 2A-2B, 3A-3B, or 4A-4B (SEQ ID NO:1, 3, 5, or 7) is intended fragments at least about 15 nt, and more preferably at least about 20 nt, still more preferably at least about 30 nt, and even more preferably, at least about 40 nt in length which are useful as diagnostic probes and primers as discussed herein. Of course larger DNA fragments 50, 100, 150, 200, 250, 300, 350, 400, 450, 500, 550, 600, 650, 700, 750, 800, 850, 900, 100, 1050, 1100, or 1115 nt in length of the sequence shown in SEQ ID NO:1 are also useful according to the present invention as are fragments corresponding to most, if not all, of the nucleotide sequence of the cDNA clone contained in the plasmid deposited as ATCC Deposit No. 97732 or as shown in SEQ ID NO:1. Similarly, larger DNA fragments 50, 100, 150, 200, 250, 300, 350, 400, 450, 500, 550, 600, 650, 700, 750, 800, 850, 900, 100, 1050, 1100, 1150, 1200, 1250, 1300, 1350,

1400, 1450, 1500, or 1525 nt in length of the sequence shown in SEQ ID NO:3 are also useful according to the present invention as are fragments corresponding to most, if not all, of the nucleotide sequence of the cDNA clone contained in the plasmid deposited as ATCC Deposit No. 97733 or as shown in SEQ ID NO:3. Similarly, larger DNA fragments 50, 100, 150, 200, 250, 300, 350, 400, 450, 500, 550, 600, 650, 700, 750, 800, 850, 900, 100, 1050, 1100, 1150, 1200, 1250, 1300, 1350, 1400, 1450, or 1464 nt in length of the sequence shown in SEQ ID NO:5 are also useful according to the present invention as are fragments corresponding to most, if not all, of the nucleotide sequence of the cDNA molecule as shown in SEQ ID NO:5. Further, larger DNA fragments 50, 100, 150, 200, 250, 300, 350, 400, 450, 500, 550, 600, 650, 700, 750, 800, 850, 900, 100, 1050, 1100, 1150, 1200, 1250, 1300, 1350, 1400, 1450, 1500, 1550, 1600, 1650, 1700, 1750, 1800, 1850, 1900, and 1908 nt in length of the sequence shown in SEQ ID NO:7 are also useful according to the present invention as are fragments corresponding to most, if not all, of the nucleotide sequence of the cDNA clone contained in the plasmid deposited as ATCC Deposit No. 97734 or as shown in SEQ ID NO:7. By a fragment at least 20 nt in length, for example, is intended fragments which include 20 or more contiguous bases from the nucleotide sequence of the deposited cDNA or the nucleotide sequence as shown in SEQ ID NOs:1, 3, 5, or 7.

Preferred nucleic acid fragments of the present invention include nucleic acid molecules encoding epitope-bearing portions of the galectin 8, 9, 10, or 10SV protein. In particular, such nucleic acid fragments of the present invention include nucleic acid molecules encoding: a polypeptide comprising amino acid residues from about 55-101, 137-162, 180-193, 216-266 in FIG. 1 (SEQ ID NO:2), 62-102, 226-259, 197-308 in FIG. 2A-2B (SEQ ID NO:4), 25-77, 84-105, 129-140, 156-183, 195-215, and 241-257 in FIG. 3A-3B (SEQ ID NO:6), and 25-77, 84-105, 129-140, and 156-183 in FIG. 4A-4B (SEQ ID NO:8). The inventors have determined that the above polypeptide fragments are antigenic regions of the galectin 8, 9, 10, and 10SV proteins. Methods for determining other such

epitope-bearing portions of the galectin 8, 9, 10, and 10SV proteins are described in detail below.

In another aspect, the invention provides an isolated nucleic acid molecule comprising a polynucleotide which hybridizes under stringent hybridization conditions to a portion of the polynucleotide in a nucleic acid molecule of the invention described above, for instance, a cDNA clone contained in ATCC Deposit Nos. 97732, 97733 and 97734. By "stringent hybridization conditions" is intended overnight incubation at 42°C in a solution comprising: 50% formamide, 5x SSC (750mN NaCl, 75mM trisodium citrate) ~~(150 mM NaCl, 15mM trisodium citrate)~~, 50 mM sodium phosphate (pH 7.6), 5x Denhardt's solution, 10% dextran sulfate, and ~~20 g/ml~~ 20 µg/ml denatured, sheared salmon sperm DNA, followed by washing the filters in 0.1x SSC at about 65°C.

By a polynucleotide which hybridizes to a "portion" of a polynucleotide is intended a polynucleotide (either DNA or RNA) hybridizing to at least about 15 nucleotides (nt), and more preferably at least about 20 nt, still more preferably at least about 30 nt, and even more preferably about 30-70 nt of the reference polynucleotide. These are useful as diagnostic probes and primers as discussed above and in more detail below.

By a portion of a polynucleotide of "at least 20 nt in length," for example, is intended 20 or more contiguous nucleotides from the nucleotide sequence of the reference polynucleotide (*e.g.*, the deposited cDNA or the nucleotide sequence as shown in FIGs. 1, 2A-2B, 3A-3B, or 4A-4B (SEQ ID NOs:1, 3, 5, or 7)). Of course, a polynucleotide which hybridizes only to a poly A sequence (such as the 3′ terminal poly(A) tract of the galectin 8, 9, 10, or 10SV cDNA shown in FIGs. 1, 2A-2B, 3A-3B, or 4A-4B, respectively (SEQ ID NOs:1, 3, 5, or 7)), or to a complementary stretch of T (or U) resides, would not be included in a polynucleotide of the invention used to hybridize to a portion of a nucleic acid of the invention, since such a polynucleotide would hybridize to any nucleic acid molecule containing a poly (A) stretch or the complement thereof (*e.g.*, practically any double-stranded cDNA clone).

As indicated, nucleic acid molecules of the present invention which encode a galectin 8, 9, 10, or 10SV polypeptide may include, but are not limited to those encoding the amino acid sequence of the polypeptide, by itself; the coding sequence for the polypeptide and additional sequences, such as those encoding an amino acid leader or secretory sequence, such as a pre-, or pro- or prepro- protein sequence; the coding sequence of the polypeptide, with or without the aforementioned additional coding sequences, together with additional, non-coding sequences, including for example, but not limited to introns and non-coding 5′ and 3′ sequences, such as the transcribed, non-translated sequences that play a role in transcription, mRNA processing, including splicing and polyadenylation signals, for example - ribosome binding and stability of mRNA; an additional coding sequence which codes for additional amino acids, such as those which provide additional functionalities. Thus, the sequence encoding the polypeptide may be fused to a marker sequence, such as a sequence encoding a peptide which facilitates purification of the fused polypeptide. In certain preferred embodiments of this aspect of the invention, the marker amino acid sequence is a hexa-histidine peptide, such as the tag provided in a pQE vector (Qiagen, Inc.), among others, many of which are commercially available. As described in Gentz *et al., Proc. Natl. Acad. Sci. USA 86*:821-824 (1989), for instance, hexa-histidine provides for convenient purification of the fusion protein. The "HA" tag is another peptide useful for purification which corresponds to an epitope derived from the influenza hemagglutinin protein, which has been described by Wilson *et al., Cell 37*:767-778 (1984). As discussed below, other such fusion proteins include the galectin 8, 9, 10, or 10SV fused to Fc at the N- or C-terminus.

The present invention further relates to variants of the nucleic acid molecules of the present invention, which encode portions, analogs or derivatives of the galectin 8, 9, 10, or 10SV protein. Variants may occur naturally, such as a natural allelic variant. By an "allelic variant" is intended one of several alternate forms of a gene occupying a given locus on a chromosome of an organism.

Genes II, Lewin, B., ed., John Wiley & Sons, New York (1985). Non-naturally occurring variants may be produced using art-known mutagenesis techniques.

Such variants include those produced by nucleotide substitutions, deletions or additions which may involve one or more nucleotides. The variants may be altered in coding regions, non-coding regions, or both. Alterations in the coding regions may produce conservative or non-conservative amino acid substitutions, deletions or additions. Especially preferred among these are silent substitutions, additions and deletions, which do not alter the properties and activities of the galectin 8, 9, 10, or 10SV protein or portions thereof. Also especially preferred in this regard are conservative substitutions.

Further embodiments of the invention include isolated nucleic acid molecules comprising a polynucleotide having a nucleotide sequence at least 95% identical, and more preferably at least 96%, 97%, 98% or 99% identical to (a) a nucleotide sequence encoding the galectin 8, 9, 10, or 10SV polypeptide having the amino acid sequence in FIGs. 1, 2A-2B, 3A-3B, or 4A-4B (SEQ ID NOs:1, 3, 5, or 7); (b) a nucleotide sequence encoding the polypeptide having the amino acid sequence in FIGs. 1, 2A-2B, 3A-3B, or 4A-4B (SEQ ID NOs:1, 3, 5, or 7), but lacking the N-terminal methionine; (c) a nucleotide sequence encoding the amino acid sequence encoded by the cDNA clone contained in ATCC Deposit Nos. 97732, 97733 or 97734 on September 24, 1996; or (d) a nucleotide sequence complementary to any of the nucleotide sequences in (a), (b), or (c).

By a polynucleotide having a nucleotide sequence at least, for example, 95% "identical" to a reference nucleotide sequence encoding a galectin 8, 9, 10, or 10SV polypeptide is intended that the nucleotide sequence of the polynucleotide is identical to the reference sequence except that the polynucleotide sequence may include up to five point mutations per each 100 nucleotides of the reference nucleotide sequence encoding the galectin 8, 9, 10, or 10SV polypeptide. In other words, to obtain a polynucleotide having a nucleotide sequence at least 95% identical to a reference nucleotide sequence, up to 5% of the nucleotides in the reference sequence may be deleted or substituted

with another nucleotide, or a number of nucleotides up to 5% of the total nucleotides in the reference sequence may be inserted into the reference sequence. These mutations of the reference sequence may occur at the 5′ or 3′ terminal positions of the reference nucleotide sequence or anywhere between those terminal positions, interspersed either individually among nucleotides in the reference sequence or in one or more contiguous groups within the reference sequence.

As a practical matter, whether any particular nucleic acid molecule is at least 95%, 96%, 97%, 98% or 99% identical to, for instance, the nucleotide sequence shown in FIGs. 1, 2A-2B, 3A-3B, or 4A-4B (SEQ ID NOs:1, 3, 5, or 7) or to the nucleotides sequence of the deposited cDNA clone can be determined conventionally using known computer programs such as the Bestfit program (Wisconsin Sequence Analysis Package, Version 8 for Unix, Genetics Computer Group, University Research Park, 575 Science Drive, Madison, WI 53711. Bestfit uses the local homology algorithm of Smith and Waterman, *Advances in Applied Mathematics 2*:482-489 (1981)), to find the best segment of homology between two sequences. When using Bestfit or any other sequence alignment program to determine whether a particular sequence is, for instance, 95% identical to a reference sequence according to the present invention, the parameters are set, of course, such that the percentage of identity is calculated over the full length of the reference nucleotide sequence and that gaps in homology of up to 5% of the total number of nucleotides in the reference sequence are allowed.

The present application is directed to nucleic acid molecules at least 95%, 96%, 97%, 98% or 99% identical to the nucleic acid sequence shown in FIGs. 1, 2A-2B, 3A-3B, or 4A-4B (SEQ ID NOs:1, 3, 5, or 7) or to the nucleic acid sequence of one of the deposited cDNAs, irrespective of whether they encode a polypeptide having galectin 8, 9, 10, or 10SV activity. This is because even where a particular nucleic acid molecule does not encode a polypeptide having galectin 8, 9, 10, or 10SV activity, one of skill in the art would still know how to

use the nucleic acid molecule, for instance, as a hybridization probe or a polymerase chain reaction (PCR) primer. Uses of the nucleic acid molecules of the present invention that do not encode a polypeptide having galectin 8, 9, 10, or 10SV activity include, *inter alia,* (1) isolating the galectin 8, 9, 10, or 10SV gene or allelic variants thereof in a cDNA library; (2) *in situ* hybridization (*e.g.*, "FISH") to metaphase chromosomal spreads to provide precise chromosomal location of the galectin 8, 9, 10, or 10SV gene, as described in Verma *et al., Human Chromosomes: A Manual of Basic Techniques*, Pergamon Press, New York (1988); and (3) Northern Blot analysis for detecting galectin 8, 9, 10, or 10SV mRNA expression in specific tissues.

Preferred, however, are nucleic acid molecules having sequences at least 95%, 96%, 97%, 98% or 99% identical to the nucleic acid sequence shown in FIGs. 1, 2A-2B, 3A-3B, or 4A-4B (SEQ ID NOs:1, 3, 5, or 7) or to the nucleic acid sequence of one of the deposited cDNAs which do, in fact, encode a polypeptide having galectin 8, 9, 10, or 10SV protein activity. By "a polypeptide having galectin 8, 9, 10, or 10SV activity" is intended polypeptides exhibiting activity similar, but not necessarily identical, to an activity of the galectin 8, 9, 10, or 10SV protein of the invention, as measured in a particular biological assay. For example, galectin 8, 9, 10, or 10SV protein activity can be measured using a lactose binding assay.

Lactose binding activity of the expressed galectin 8, 9, 10, or 10SV is assayed by immunodetection of *in situ* binding activity to asialofetuin (Sigma) immobilized on nitrocellulose (Amersham) (Madsen *et al., J. Biol. Chem. 270(11):*5823-5829 (1995)). Thirty μg of asialofetuin dissolved in 3 μl of water is spotted on a 1-cm^2 strip of nitrocellulose. The nitrocellulose pieces are then placed in a 24-well tissue culture plate and incubated overnight in buffer B (58 mM Na_2HPO_4, 18 mM KH_2PO_4, 75 mM NaCl, 2 mM EDTA, and 3% BSA, pH7.2) with constant agitation at 22°C. Following incubation, the blocking medium is aspirated and the nitrocellulose pieces are washed three times in buffer A (58 mM Na_2HPO_4, 18 mM KH_2PO_4, 75 mM NaCl, 2 mM EDTA, 4 mM β-

mercaptoethanol and 0.2% BSA, pH7.2). Cell extracts (preferably, COS cells) are prepared containing 1% BSA and either with or without 150 mM lactose (105 µl of primary extract, 15 µl of 10% BSA in buffer A and either 30 µl of 0.75 M lactose in buffer A or 30 µl of buffer A). The immobilized asialofetuin is incubated with the extracts for 2 h and washed 5 times in buffer A. The nitrocellulose pieces are then fixed in 2% formalin in PBS (58 mM Na_2HPO_4, 18 mM KH_2PO_4, 75 mM NaCl, 2 mM EDTA pH7.2) for 1 hour to prevent loss of bound galectin. Following extensive washing in PBS the pieces were incubated with rabbit anti-galectin 8, 9, 10, or 10SV polyclonal serum diluted 1:100 in PBS for 2 h at 22°C. The pieces are then washed in PBS and incubated with peroxidase-labeled goat anti-rabbit antibodies (DAKO). Following incubation for 2 h at 22°C, the pieces are washed in PBS and the substrate is added. Nitrocellulose pieces are incubated until the color develops and the reaction is stopped by washing in distilled water.

Of course, due to the degeneracy of the genetic code, one of ordinary skill in the art will immediately recognize that a large number of the nucleic acid molecules having a sequence at least 95%, 96%, 97%, 98%, or 99% identical to the nucleic acid sequence of the deposited cDNA or the nucleic acid sequence shown in FIGs. 1, 2A-2B, 3A-3B, or 4A-4B (SEQ ID NOs:1, 3, 5, or 7, respectively) will encode "a polypeptide having galectin 8, 9, 10, or 10SV protein activity." In fact, since degenerate variants of these nucleotide sequences all encode the same polypeptide, this will be clear to the skilled artisan even without performing the above described comparison assay. It will be further recognized in the art that, for such nucleic acid molecules that are not degenerate variants, a reasonable number will also encode a polypeptide having galectin 8, 9, 10, or 10SV protein activity. This is because the skilled artisan is fully aware of amino acid substitutions that are either less likely or not likely to significantly effect protein function (*e.g.*, replacing one aliphatic amino acid with a second aliphatic amino acid).

For example, guidance concerning how to make phenotypically silent amino acid substitutions is provided in Bowie, J. U. *et al.,* "Deciphering the Message in Protein Sequences: Tolerance to Amino Acid Substitutions," *Science 247:*1306-1310 (1990), wherein the authors indicate that proteins are surprisingly tolerant of amino acid substitutions.

Vectors and Host Cells

The present invention also relates to vectors which include the isolated DNA molecules of the present invention, host cells which are genetically engineered with the recombinant vectors, and the production of galectin 8, 9, 10, or 10SV polypeptides or fragments thereof by recombinant techniques.

The polynucleotides may be joined to a vector containing a selectable marker for propagation in a host. Generally, a plasmid vector is introduced in a precipitate, such as a calcium phosphate precipitate, or in a complex with a charged lipid. If the vector is a virus, it may be packaged *in vitro* using an appropriate packaging cell line and then transduced into host cells.

The DNA insert should be operatively linked to an appropriate promoter, such as the phage lambda PL promoter, the *E. coli lac, trp* and *tac* promoters, the SV40 early and late promoters and promoters of retroviral LTRs, to name a few. Other suitable promoters will be known to the skilled artisan. The expression constructs will further contain sites for transcription initiation, termination and, in the transcribed region, a ribosome binding site for translation. The coding portion of the transcripts expressed by the constructs will preferably include a translation initiating at the beginning and a termination codon (UAA, UGA or UAG) appropriately positioned at the end of the polypeptide to be translated.

As indicated, the expression vectors will preferably include at least one selectable marker. Such markers include dihydrofolate reductase or neomycin resistance for eukaryotic cell culture and tetracycline or ampicillin resistance genes for culturing in *E. coli* and other bacteria. Representative examples of

appropriate hosts include, but are not limited to, bacterial cells, such as *E. coli*, *Streptomyces* and *Salmonella typhimurium* cells; fungal cells, such as yeast cells; insect cells such as *Drosophila* S2 and *Spodoptera* Sf9 cells; animal cells such as CHO, COS and Bowes melanoma cells; and plant cells. Appropriate culture mediums and conditions for the above-described host cells are known in the art.

Among vectors preferred for use in bacteria include pQE70, pQE60 and pQE-9, available from Qiagen; pBS vectors, Phagescript vectors, Bluescript vectors, pNH8A, pNH16a, pNH18A, pNH46A, available from Stratagene; and ptrc99a, pKK223-3, pKK233-3, pDR540, pRIT5 available from Pharmacia. Among preferred eukaryotic vectors are pWLNEO, pSV2CAT, pOG44, pXT1 and pSG available from Stratagene; and pSVK3, pBPV, pMSG and pSVL available from Pharmacia. Other suitable vectors will be readily apparent to the skilled artisan.

Introduction of the construct into the host cell can be effected by calcium phosphate transfection, DEAE-dextran mediated transfection, cationic lipid-mediated transfection, electroporation, transduction, infection or other methods. Such methods are described in many standard laboratory manuals, such as Davis *et al., Basic Methods In Molecular Biology* (1986).

The polypeptide may be expressed in a modified form, such as a fusion protein, and may include not only secretion signals, but also additional heterologous functional regions. For instance, a region of additional amino acids, particularly charged amino acids, may be added to the N-terminus of the polypeptide to improve stability and persistence in the host cell, during purification, or during subsequent handling and storage. Also, peptide moieties may be added to the polypeptide to facilitate purification. Such regions may be removed prior to final preparation of the polypeptide. The addition of peptide moieties to polypeptides to engender secretion or excretion, to improve stability and to facilitate purification, among others, are familiar and routine techniques in the art. A preferred fusion protein comprises a heterologous region from immunoglobulin that is useful to solubilize proteins. For example, EP-A-O 464

533 (Canadian counterpart 2045869) discloses fusion proteins comprising various portions of constant region of immunoglobin molecules together with another human protein or part thereof. In many cases, the Fc part in a fusion protein is thoroughly advantageous for use in therapy and diagnosis and thus results, for example, in improved pharmacokinetic properties (EP-A 0232 262). On the other hand, for some uses it would be desirable to be able to delete the Fc part after the fusion protein has been expressed, detected and purified in the advantageous manner described. This is the case when Fc portion proves to be a hindrance to use in therapy and diagnosis, for example when the fusion protein is to be used as antigen for immunizations. In drug discovery, for example, human proteins, such as, hIL5-receptor has been fused with Fc portions for the purpose of high-throughput screening assays to identify antagonists of hIL-5. See, D. Bennett *et al.*, *Journal of Molecular Recognition*, Vol. 8 52-58 (1995) and K. Johanson *et al.*, *The Journal of Biological Chemistry*, Vol. 270, No. 16, pp 9459-9471 (1995).

The galectin 8, 9, 10, or 10SV protein can be recovered and purified from recombinant cell cultures by well-known methods including ammonium sulfate or ethanol precipitation, acid extraction, anion or cation exchange chromatography, phosphocellulose chromatography, hydrophobic interaction chromatography, affinity chromatography, hydroxylapatite chromatography and lectin chromatography. Most preferably, high performance liquid chromatography ("HPLC") is employed for purification. Polypeptides of the present invention include naturally purified products, products of chemical synthetic procedures, and products produced by recombinant techniques from a prokaryotic or eukaryotic host, including, for example, bacterial, yeast, higher plant, insect and mammalian cells. Depending upon the host employed in a recombinant production procedure, the polypeptides of the present invention may be glycosylated or may be non-glycosylated. In addition, polypeptides of the invention may also include an initial modified methionine residue, in some cases as a result of host-mediated processes.

A 8, 9, 10 and 10SV Polypeptides

Galectin ~~8, 9, and 10 Polypeptides~~ and Fragments

The invention further provides an isolated galectin 8, 9, 10, or 10SV polypeptide having (1) the amino acid sequence encoded by one of the deposited cDNAs, (2) the amino acid sequence in FIGs. 1, 2A-2B, 3A-3B, or 4A-4B (SEQ ID NOs:2, 4, 6, or 8, respectively), or (3) the amino acid sequence of a peptide or polypeptide comprising a portion of the above polypeptides.

It will be recognized in the art that some amino acid sequences of the galectin 8, 9, 10, or 10SV polypeptide can be varied without significant effect of the structure or function of the protein. If such differences in sequence are contemplated, it should be remembered that there will be critical areas on the protein which determine activity.

Thus, the invention further includes variations of the galectin 8, 9, 10, or 10SV polypeptide which show substantial galectin 8, 9, 10, or 10SV polypeptide activity or which include regions of galectin 8, 9, 10, or 10SV protein such as the protein portions discussed below. Such mutants include deletions, insertions, inversions, repeats, and type substitutions. As indicated above, guidance concerning which amino acid changes are likely to be phenotypically silent can be found in Bowie, J.U., *et al.*, "Deciphering the Message in Protein Sequences: Tolerance to Amino Acid Substitutions," *Science 247:*1306-1310 (1990).

Thus, the fragment, derivative or analog of the polypeptide of SEQ ID NOs:2, 4, 6, or 8, or that encoded by one of the deposited cDNAs, may be (i) one in which one or more of the amino acid residues are substituted with a conserved or non-conserved amino acid residue (preferably a conserved amino acid residue) and such substituted amino acid residue may or may not be one encoded by the genetic code, or (ii) one in which one or more of the amino acid residues includes a substituent group, or (iii) one in which the mature polypeptide is fused with another compound, such as a compound to increase the half-life of the polypeptide (for example, polyethylene glycol), or (iv) one in which the additional amino acids are fused to the mature polypeptide, such as an IgG Fc

fusion region peptide or leader or secretory sequence or a sequence which is employed for purification of the mature polypeptide or a proprotein sequence. Such fragments, derivatives and analogs are deemed to be within the scope of those skilled in the art from the teachings herein.

Of particular interest are substitutions of charged amino acids with another charged amino acid and with neutral or negatively charged amino acids. The latter results in proteins with reduced positive charge to improve the characteristics of a galectin 8, 9, 10, or 10SV protein. The prevention of aggregation is highly desirable. Aggregation of proteins not only results in a loss of activity but can also be problematic when preparing pharmaceutical formulations, because they can be immunogenic. (Pinckard *et al., Clin. Exp. Immunol. 2*:331-340 (1967); Robbins *et al., Diabetes 36*:838-845 (1987); Cleland *et al., Crit. Rev. Therapeutic Drug Carrier Systems 10*:307-377 (1993)).

As indicated, changes are preferably of a minor nature, such as conservative amino acid substitutions that do not significantly affect the folding or activity of the protein (see Table 1).

TABLE 1. Conservative Amino Acid Substitutions.

Aromatic	Phenylalanine Tryptophan Tyrosine
Hydrophobic	Leucine Isoleucine Valine
Polar	Glutamine Asparagine
Basic	Arginine Lysine Histidine
Acidic	Aspartic Acid Glutamic Acid
Small	Alanine Serine Threonine Methionine Glycine

Of course, the number of amino acid substitutions a skilled artisan would make depends on many factors, including those described above and below. Generally speaking, the number of substitutions for any given galectin 8, 9, 10, or 10SV polypeptide or mutant thereof will not be more than 50, 40, 30, 20, 10, 5, or 3, depending on the objective.

Amino acids in a galectin 8, 9, 10, or 10SV protein of the present invention that are essential for function can be identified by methods known in the art, such as site-directed mutagenesis or alanine-scanning mutagenesis (Cunningham and Wells, *Science 244*:1081-1085 (1989)). The latter procedure introduces single alanine mutations at every residue in the molecule. Sites that are critical for ligand binding can also be determined by structural analysis such as crystallization, nuclear magnetic resonance or photoaffinity labeling (Smith *et al., J. Mol. Biol. 224*:899-904 (1992) and de Vos *et al., Science 255*:306-312 (1992)).

The polypeptides of the present invention are preferably provided in an isolated form. By "isolated polypeptide" is intended a polypeptide removed from its native environment. Thus, a polypeptide produced and/or contained within a recombinant host cell is considered isolated for purposes of the present invention. Also intended as an "isolated polypeptide" are polypeptides that have been purified, partially or substantially, from a recombinant host cell. For example, a recombinantly produced version of a galectin 8, 9, 10, or 10SV polypeptide can be substantially purified by the one-step method described in Smith and Johnson, *Gene 67:*31-40 (1988).

The polypeptides of the present invention include the polypeptides encoded by the deposited cDNAs; a polypeptide comprising amino acids about 1 to about 323 in SEQ ID NO:2, about 1 to about 311 in SEQ ID NO:4, about 1 to about 317 in SEQ ID NO:6, and about 1 to about 200 in SEQ ID NO:8; a polypeptide comprising amino acids about 2 to about 323 in SEQ ID NO:2, about 2 to about 311 in SEQ ID NO:4, about 2 to about 317 in SEQ ID NO:6 and about 2 to about 200 in SEQ ID NO:8; as well as polypeptides which are at least 95% identical, still more preferably at least 96%, 97%, 98% or 99% identical to the polypeptides described above and also include portions of such polypeptides with at least 30 amino acids and more preferably at least 50 amino acids.

By a polypeptide having an amino acid sequence at least, for example, 95% "identical" to a reference amino acid sequence of a galectin 8, 9, 10, or 10SV polypeptide is intended that the amino acid sequence of the polypeptide is identical to the reference sequence except that the polypeptide sequence may include up to five amino acid alterations per each 100 amino acids of the reference amino acid of the galectin 8, 9, 10, or 10SV polypeptide. In other words, to obtain a polypeptide having an amino acid sequence at least 95% identical to a reference amino acid sequence, up to 5% of the amino acid residues in the reference sequence may be deleted or substituted with another amino acid, or a number of amino acids up to 5% of the total amino acid residues in the reference sequence may be inserted into the reference sequence. These alterations

of the reference sequence may occur at the amino or carboxy terminal positions of the reference amino acid sequence or anywhere between those terminal positions, interspersed either individually among residues in the reference sequence or in one or more contiguous groups within the reference sequence.

As a practical matter, whether any particular polypeptide is at least 95%, 96%, 97%, 98% or 99% identical to, for instance, the amino acid sequence shown in FIGs. 1, 2A-2B, 3A-3B, or 4A-4B (SEQ ID NOs:2, 4, 6, or 8, respectively) or to the amino acid sequence encoded by one of the deposited cDNA clones (ATCC Deposit Numbers 97732, 97733 and 97734) can be determined conventionally using known computer programs such the Bestfit program (Wisconsin Sequence Analysis Package, Version 8 for Unix, Genetics Computer Group, University Research Park, 575 Science Drive, Madison, WI 53711). When using Bestfit or any other sequence alignment program to determine whether a particular sequence is, for instance, 95% identical to a reference sequence according to the present invention, the parameters are set, of course, such that the percentage of identity is calculated over the full length of the reference amino acid sequence and that gaps in homology of up to 5% of the total number of amino acid residues in the reference sequence are allowed.

The polypeptide of the present invention could be used as a molecular weight marker on SDS-PAGE gels or on molecular sieve gel filtration columns using methods well known to those of skill in the art.

In another aspect, the invention provides a peptide or polypeptide comprising an epitope-bearing portion of a polypeptide of the invention. The epitope of this polypeptide portion is an immunogenic or antigenic epitope described herein. An "immunogenic epitope" is defined as a part of a protein that elicits an antibody response when the whole protein is the immunogen. On the other hand, a region of a protein molecule to which an antibody can bind is defined as an "antigenic epitope." The number of immunogenic epitopes of a protein generally is less than the number of antigenic epitopes. See, for instance, Geysen *et al., Proc. Natl. Acad. Sci. USA 81*:3998- 4002 (1983).

As to the selection of peptides or polypeptides bearing an antigenic epitope (*i.e.*, that contain a region of a protein molecule to which an antibody can bind), it is well known in that art that relatively short synthetic peptides that mimic part of a protein sequence are routinely capable of eliciting an antiserum that reacts with the partially mimicked protein. See, for instance, Sutcliffe, J. G., Shinnick, T. M., Green, N. and Learner, R.A. (1983) Antibodies that react with predetermined sites on proteins. *Science 219:*660-666. Peptides capable of eliciting protein-reactive sera are frequently represented in the primary sequence of a protein, can be characterized by a set of simple chemical rules, and are confined neither to immunodominant regions of intact proteins (*i.e.*, immunogenic epitopes) nor to the amino or carboxyl terminals.

Antigenic epitope-bearing peptides and polypeptides of the invention are therefore useful to raise antibodies, including monoclonal antibodies, that bind specifically to a polypeptide of the invention. See, for instance, Wilson *et al.*, *Cell 37*:767-778 (1984) at 777.

Antigenic epitope-bearing peptides and polypeptides of the invention preferably contain a sequence of at least seven, more preferably at least nine and most preferably between about at least 15 to about 30 amino acids contained within the amino acid sequence of a polypeptide of the invention.

Non-limiting examples of antigenic polypeptides or peptides that can be used to generate galectin 8, 9, 10, or 10SV-specific antibodies include: a polypeptide comprising amino acid residues from about 55-101, 137-162, 180-193, 216-266 in FIG. 1 (SEQ ID NO:2), 62-102, 226-259, 197-308 in FIG. 2A-2B (SEQ ID NO:4), 25-77, 84-105, 129-140, 156-183, 195-215, and 241-257 in FIG. 3A-3B (SEQ ID NO:6), and 25-77, 84-105, 129-140, and 156-183 in FIG. 4A-4B (SEQ ID NO:8), respectively. As indicated above, the inventors have determined that the above polypeptide fragments are antigenic regions of the galectin 8, 9, 10, or 10SV protein.

The epitope-bearing peptides and polypeptides of the invention may be produced by any conventional means Houghten, R. A. (1985) General method

for the rapid solid-phase synthesis of large numbers of peptides: specificity of antigen-antibody interaction at the level of individual amino acids. *Proc. Natl. Acad. Sci. USA 82*:5131-5135. This "Simultaneous Multiple Peptide Synthesis (SMPS)" process is further described in U.S. Patent No. 4,631,211 to Houghten *et al.* (1986).

As one of skill in the art will appreciate, galectin 8, 9, 10, or 10SV polypeptides of the present invention and the epitope-bearing fragments thereof described above can be combined with parts of the constant domain of immunoglobulins (IgG), resulting in chimeric polypeptides. These fusion proteins facilitate purification and show an increased half-life *in vivo*. This has been shown, *e.g.*, for chimeric proteins consisting of the first two domains of the human CD4-polypeptide and various domains of the constant regions of the heavy or light chains of mammalian immunoglobulins (EPA 394,827; Traunecker *et al., Nature 331*:84- 86 (1988)). Fusion proteins that have a disulfide-linked dimeric structure due to the IgG part can also be more efficient in binding and neutralizing other molecules than the monomeric galectin 8, 9, 10, or 10SV protein or protein fragment alone (Fountoulakis *et al., J Biochem 270*:3958-3964 (1995)).

Diagnosis and Prognosis

It is believed that certain tissues in mammals with certain diseases (cancer, autoimmune diseases, inflammatory diseases, asthma, and allergic diseases) express significantly altered (enhanced or decreased) levels of the galectin 8, 9, 10, or 10SV protein and mRNA encoding the galectin 8, 9, 10, or 10SV protein when compared to a corresponding "standard" mammal, *i.e.*, a mammal of the same species not having the disease. Further, it is believed that altered levels of the galectin 8, 9, 10, or 10SV protein can be detected in certain body fluids (*e.g.*, sera, plasma, urine, and spinal fluid) from mammals with the disease when compared to sera from mammals of the same species not having the

disease. Thus, the invention provides a diagnostic method useful during diagnosis, which involves assaying the expression level of the gene encoding the galectin 8, 9, 10, or 10SV protein in mammalian cells or body fluid and comparing the gene expression level with a standard galectin 8, 9, 10, or 10SV gene expression level, whereby an increase or decrease in the gene expression level over the standard is indicative of the disease.

Where a diagnosis has already been made according to conventional methods, the present invention is useful as a prognostic indicator, whereby patients exhibiting altered galectin 8, 9, 10, or 10SV gene expression will experience a worse clinical outcome relative to patients expressing the gene at a normal level.

By "assaying the expression level of the gene encoding the galectin 8, 9, 10, or 10SV protein" is intended qualitatively or quantitatively measuring or estimating the level of the galectin 8, 9, 10, or 10SV protein or the level of the mRNA encoding the galectin 8, 9, 10, or 10SV protein in a first biological sample either directly (*e.g.*, by determining or estimating absolute protein level or mRNA level) or relatively (*e.g.*, by comparing to the galectin 8, 9, 10, or 10SV protein level or mRNA level in a second biological sample).

Preferably, the galectin 8, 9, 10, or 10SV protein level or mRNA level in the first biological sample is measured or estimated and compared to a standard galectin 8, 9, 10, or 10SV protein level or mRNA level, the standard being taken from a second biological sample obtained from an individual not having the cancer. As will be appreciated in the art, once a standard galectin 8, 9, 10, or 10SV protein level or mRNA level is known, it can be used repeatedly as a standard for comparison.

By "biological sample" is intended any biological sample obtained from an individual, cell line, tissue culture, or other source which contains galectin 8, 9, 10, or 10SV protein or mRNA. Biological samples include mammalian body fluids (such as sera, plasma, urine, synovial fluid and spinal fluid) which contain

secreted galectin 8, 9, 10, or 10SV protein, and ovarian, prostate, heart, placenta, pancreas liver, spleen, lung, breast and umbilical tissue.

The present invention is useful for detecting diseases in mammals (for example, cancer, autoimmune diseases, inflammatory diseases, asthma, and allergic diseases). In particular the invention is useful during diagnosis of the of following types of cancers in mammals: melanoma, renal astrocytoma, Hodgkin disease, breast, ovarian, prostate, bone, liver, lung, pancreatic, and spleenic. Preferred mammals include monkeys, apes, cats, dogs, cows, pigs, horses, rabbits and humans. Particularly preferred are humans.

Total cellular RNA can be isolated from a biological sample using the single-step guanidinium-thiocyanate-phenol-chloroform method described in Chomczynski and Sacchi, *Anal. Biochem. 162:*156-159 (1987). Levels of mRNA encoding the galectin 8, 9, 10, or 10SV protein are then assayed using any appropriate method. These include Northern blot analysis, (Harada *et al.*, *Cell 63:*303-312 (1990) S1 nuclease mapping, (Fijita *et al., Cell 49:*357-367 (1987)) the polymerase chain reaction (PCR), reverse transcription in combination with the polymerase chain reaction (RT-PCR) (Makino *et al.*, *Technique 2:*295-301 (1990), and reverse transcription in combination with the ligase chain reaction (RT-LCR).

Assaying galectin 8, 9, 10, or 10SV protein levels in a biological sample can be performed via antibody-based techniques. For example, galectin 8, 9, 10, or 10SV protein expression in tissues can be studied with classical immunohistological methods. (Jalkanen, M., *et al., J. Cell. Biol. 101:*976-985 (1985); Jalkanen, M., *et al., J. Cell . Biol. 105:*3087-3096 (1987)).

Other antibody-based methods useful for detecting galectin 8, 9, 10, or 10SV protein gene expression include immunoassays, such as the enzyme linked immunosorbent assay (ELISA) and the radioimmunoassay (RIA).

Suitable labels are known in the art and include enzyme labels, such as, Glucose oxidase, and radioisotopes, such as iodine (^{125}I, ^{121}I), carbon (^{14}C), sulfur

(^{35}S), tritium (^{3}H), indium (^{112}In), and technetium (^{99m}Tc), and fluorescent labels, such as fluorescein and rhodamine, and biotin.

Therapeutics

It is to be understood that although the following discussion is specifically directed to human patients, the teachings are also applicable to any animal that expresses galectin 8, 9, 10, or 10SV.

As noted above, galectin 8, 9, 10, and 10SV share significant homology with other galectins. Galectin 1 induces apoptosis of T cells and T cell leukemia cell lines. Thus, it is believed by the inventors that galectin 8, 9, 10, and 10SV are active in modulating growth regulatory activities, immunomodulatory activity, cell-cell and cell-substrate interactions, and apoptosis.

The ability of galectin 8, 9, 10, or 10SV to modulate growth regulatory activity may be therapeutically valuable in the treatment of clinical manifestations of such cell regulatory disorders. Disorders which can be treated include, but should not be limited to, autoimmune disease, cancer (preferably, melanoma, renal, astrocytoma, and Hodgkin disease), inflammatory disease, wound healing, arteriosclerosis, other heart diseases, microbe infection (virus, fungal, bacterial, and parasite), asthma, and allergic diseases.

Given the activities modulated by galectin 8, 9, 10, and 10SV, it is readily apparent that a substantially altered (increased or decreased) level of expression of galectin 8, 9, 10, or 10SV in an individual compared to the standard or "normal" level produces pathological conditions such as those described above. It will also be appreciated by one of ordinary skill that the galectin 8, 9, 10, or 10SV protein of the invention will exert its modulating activities on any of its target cells. Therefore, it will be appreciated that conditions caused by a decrease in the standard or normal level of galectin 8, 9, 10, or 10SV activity in an individual, can be treated by administration of galectin 8, 9, 10, or 10SV protein or an agonist thereof. Thus, the invention further provides a method of treating

an individual in need of an increased level of galectin 8, 9, 10, or 10SV activity comprising administering to such an individual a pharmaceutical composition comprising an amount of an isolated galectin 8, 9, 10, or 10SV polypeptide of the invention or an agonist thereof to increase the galectin 8, 9, 10, or 10SV activity level in such an individual.

A still further aspect of the invention is related to a method for treating an individual in need of a decreased level of galectin 8, 9, 10, or 10SV activity in the body comprising, administering to such an individual a composition comprising a therapeutically effective amount of a galectin 8, 9, 10, or 10SV antagonist. Preferred antagonists for use in the present invention are galectin 8, 9, 10, or 10SV-specific antibodies.

Modes of administration

It will be appreciated that conditions caused by a decrease in the standard or normal level of galectin 8, 9, 10, or 10SV activity in an individual, can be treated by administration of galectin 8, 9, 10, or 10SV protein or an agonist thereof. Thus, the invention further provides a method of treating an individual in need of an increased level of galectin 8, 9, 10, or 10SV activity comprising administering to such an individual a pharmaceutical composition comprising an effective amount of an isolated galectin 8, 9, 10, or 10SV polypeptide of the invention, particularly a mature form of the galectin 8, 9, 10, or 10SV, effective to increase the galectin 8, 9, 10, or 10SV activity level in such an individual.

As a general proposition, the total pharmaceutically effective amount of galectin 8, 9, 10, or 10SV polypeptide administered parenterally per dose will be in the range of about 1 μg/kg/day to 10 mg/kg/day of patient body weight, although, as noted above, this will be subject to therapeutic discretion. More preferably, this dose is at least 0.01 mg/kg/day, and most preferably for humans between about 0.01 and 1 mg/kg/day for the hormone. If given continuously, the galectin 8, 9, 10, or 10SV polypeptide is typically administered at a dose rate of

about 1 μg/kg/hour to about 50 μg/kg/hour, either by 1-4 injections per day or by continuous subcutaneous infusions, for example, using a mini-pump. An intravenous bag solution may also be employed.

Pharmaceutical compositions containing the galectin 8, 9, 10, or 10SV of the invention may be administered orally, rectally, parenterally, intracistemally, intravaginally, intraperitoneally, topically (as by powders, ointments, drops or transdermal patch), bucally, or as an oral or nasal spray. By "pharmaceutically acceptable carrier" is meant a non-toxic solid, semisolid or liquid filler, diluent, encapsulating material or formulation auxiliary of any type. The term "parenteral" as used herein refers to modes of administration which include intravenous, intramuscular, intraperitoneal, intrasternal, subcutaneous and intraarticular injection and infusion.

Chromosome Assays

The nucleic acid molecules of the present invention are also valuable for chromosome identification. The sequence is specifically targeted to and can hybridize with a particular location on an individual human chromosome. The mapping of DNAs to chromosomes according to the present invention is an important first step in correlating those sequences with genes associated with disease.

In certain preferred embodiments in this regard, the cDNA herein disclosed is used to clone genomic DNA of a galectin 8, 9, 10, or 10SV protein gene. This can be accomplished using a variety of well known techniques and libraries, which generally are available commercially. The genomic DNA then is used for *in situ* chromosome mapping using well known techniques for this purpose.

In addition, in some cases, sequences can be mapped to chromosomes by preparing PCR primers (preferably 15-25 bp) from the cDNA. Computer analysis of the 3′ untranslated region of the gene is used to rapidly select primers that do

not span more than one exon in the genomic DNA, thus complicating the amplification process. These primers are then used for PCR screening of somatic cell hybrids containing individual human chromosomes.

Fluorescence *in situ* hybridization ("FISH") of a cDNA clone to a metaphase chromosomal spread can be used to provide a precise chromosomal location in one step. This technique can be used with probes from the cDNA as short as 50 or 60 bp. For a review of this technique, see Verma *et al., Human Chromosomes: A Manual Of Basic Techniques*, Pergamon Press, New York (1988).

Once a sequence has been mapped to a precise chromosomal location, the physical position of the sequence on the chromosome can be correlated with genetic map data. Such data are found, for example, in V. McKusick, *Mendelian Inheritance In Man*, available on-line through Johns Hopkins University, Welch Medical Library. The relationship between genes and diseases that have been mapped to the same chromosomal region are then identified through linkage analysis (coinheritance of physically adjacent genes).

Next, it is necessary to determine the differences in the cDNA or genomic sequence between affected and unaffected individuals. If a mutation is observed in some or all of the affected individuals but not in any normal individuals, then the mutation is likely to be the causative agent of the disease.

Having generally described the invention, the same will be more readily understood by reference to the following examples, which are provided by way of illustration and are not intended as limiting.

Examples

Example 1: Expression and Purification of Galectin 8, 9, 10 and 10SV in E. coli

The DNA sequence encoding the galectin 9 protein in the deposited cDNA clone was amplified using PCR oligonucleotide primers specific to the amino terminal sequences of the galectin 9 protein and to vector sequences 3′ to

the gene. Additional nucleotides containing restriction sites to facilitate cloning are added to the 5′ and 3′ sequences.

The DNA sequence encoding the galectin 8 or 10SV protein in the deposited cDNA clone is amplified using PCR oligonucleotide primers specific to the nucleotide sequences encoding the amino terminal sequences of the galectin 8 or 10SV protein and to vector sequences 3′ to the gene. Additional nucleotides containing restriction sites to facilitate cloning are added to the 5′ and 3′ sequences.

The cDNA sequence encoding the galectin 10 protein is amplified from either a human endometrial tumor or human fetal heart cDNA library using PCR oligonucleotide primers specific to the nucleotide sequences encoding the amino terminal sequences of the galectin 10 protein and to vector sequences 3′ to the gene. Additional nucleotides containing restriction sites to facilitate cloning are added to the 5′ and 3′ sequences.

The 5′ galectin 8 oligonucleotide primer has the sequence 5' cgc ccATGg CCTATGTCCCCGCACCG 3' (SEQ ID NO:41) containing the underlined NcoI restriction site and nucleotides 56 to 72 of the galectin 8 protein coding sequence in FIG. 1 (SEQ ID NO:1).

The 3′ galectin 8 primer has the sequence 5′ cgc AAG CTT TTAGATC TGGACATAGGAC 3′ (SEQ ID NO:42) containing the underlined HindIII restriction site followed by nucleotides complementary to position 1005 to 1023 of the galectin 8 protein coding sequence in FIG. 1 (SEQ ID NO:1).

The 5′ galectin 9 oligonucleotide primer has the sequence 5′cgc ccATGg CCTT CAGCGGTTCCCAG 3′ (SEQ ID NO:43) containing the underlined NcoI restriction site and nucleotides 20 to 36 of the galectin 9 protein coding sequence in FIG. 2A-2B (SEQ ID NO:3).

The 3′ galectin 9 primer has the sequence 5′cgc AAG CTT CAGGGTT GGAAAGGCTG (SEQ ID NO:44) containing the underlined HindIII restriction site followed by nucleotides complementary to position 1029 to 1045 of the galectin 9 protein coding sequence in FIG. 2A-2B (SEQ ID NO:3).

The 5′ galectin 10 and 10SV oligonucleotide primer has the sequence 5′cgc CCATGc TGTTGTCCTTAAACAAC 3′ (SEQ ID NO:45) containing the underlined SphI restriction site and nucleotides 122-138 of the galectin 10 protein coding sequence in FIG. 3A-3B (SEQ ID NO:5).

The 3′ galectin 10 primer has the sequence 5' cgc CTG CAG CACAGAA GCCATTCTG 3′ (SEQ ID NO:46) containing the underlined PstI restriction site followed by nucleotides complementary to position 1105-1120 of the galectin 10 protein coding sequence in FIG. 3A-3B (SEQ ID NO:5).

The 3′ galectin 10SV primer has the sequence 5' CGCCTGCAGCTA TGCAACTTTATAAAATATTCC 3′ (SEQ ID NO:47) containing the underlined PstI restriction site followed by nucleotides complementary to 3' end of the galectin 10SV protein coding sequence in FIG. 4A-4B (SEQ ID NO:7).

The restriction sites are convenient to restriction enzyme sites in the bacterial expression vector pQE60 (galectin 8 and 9) or pQE6 (galectin 10), which are used for bacterial expression in these examples. (Qiagen, Inc. 9259 Eton Avenue, Chatsworth, CA, 91311). pQE60 encodes ampicillin antibiotic resistance ("Amp'") and contains a bacterial origin of replication ("ori"), an IPTG inducible promoter, a ribosome binding site ("RBS"), a 6-His tag and restriction enzyme sites.

The amplified galectin 8, 9, 10, or 10SV DNA and the vector pQE60 or pQE6 both are digested with NcoI and HindIII (for galectin 8 and 9) or SphI and PstI (for galectin 10) and the digested DNAs are then ligated together. Insertion of the galectin 8, 9, 10, or 10SV protein DNA into the restricted pQE60 or pQE6 vector placed the galectin 8, 9, 10, or 10SV protein coding region downstream of and operably linked to the vector's IPTG-inducible promoter and in-frame with an initiating AUG appropriately positioned for translation of galectin 8, 9, 10, or 10SV protein.

The ligation mixture is transformed into competent *E. coli* cells using standard procedures. Such procedures are described in Sambrook *et al.*, MOLECULAR CLONING: A LABORATORY MANUAL, 2nd Ed.; Cold Spring

Harbor Laboratory Press, Cold Spring Harbor, N.Y. (1989). *E. coli* strain M15/rep4, containing multiple copies of the plasmid pREP4, which expresses lac repressor and confers kanamycin resistance ("Kan^r"), is used in carrying out the example described herein. This strain, which is only one of many that are suitable for expressing galectin 8, 9, 10, or 10SV protein, is available commercially from Qiagen.

Transformants are identified by their ability to grow on LB plates in the presence of ampicillin and kanamycin. Plasmid DNA is isolated from resistant colonies and the identity of the cloned DNA confirmed by restriction analysis.

Clones containing the desired constructs are grown overnight ("O/N") in liquid culture in LB media supplemented with both ampicillin (100 μg/ml) and kanamycin (25 μg/ml).

The O/N culture is used to inoculate a large culture, at a dilution of approximately 1:100 to 1:250. The cells are grown to an optical density at 600nm ("OD600") of between 0.4 and 0.6. Isopropyl-B-D-thiogalactopyranoside ("IPTG") is then added to a final concentration of 1 mM to induce transcription from *lac* repressor sensitive promoters, by inactivating the *lac*I repressor. Cells subsequently are incubated further for 3 to 4 hours. Cells then are harvested by centrifugation and disrupted, by standard methods. Inclusion bodies are purified from the disrupted cells using routine collection techniques, and protein is solubilized from the inclusion bodies into 8M urea. The 8M urea solution containing the solubilized protein is passed over a PD-10 column in 2X phosphate-buffered saline ("PBS"), thereby removing the urea, exchanging the buffer and refolding the protein. The protein is purified by a further step of chromatography to remove endotoxin. Then, it is sterile filtered. The sterile filtered protein preparation is stored in 2X PBS at a concentration of 95 μ/ml.

Example 2: Cloning and Expression of Galectin 8, 9, 10 and 10SV protein in a Baculovirus Expression System

The cDNA sequence encoding the full length galectin 8, 9, 10, or 10SV protein in the deposited clone is amplified using PCR oligonucleotide primers corresponding to the 5′ and 3′ sequences of the gene:

The 5′ galectin 8 oligonucleotide primer has the sequence 5′cgc CCC GGG GCCTATGTCCCCGCAC 3′ (SEQ ID NO:48) containing the underlined SmaI restriction site and nucleotides 55 to 70 of the galectin 8 protein coding sequence in FIG. 1 (SEQ ID NO:1).

The 3′ galectin 8 primer has the sequence 5′ cgc GGT ACC TTAGATCTGG ACATAGGAC 3′ (SEQ ID NO:49) containing the underlined Asp718 restriction site followed by nucleotides complementary to position 1005 to 1023 of the galectin 8 protein coding sequence in FIG. 1 (SEQ ID NO:1).

The 5′ galectin 9 oligonucleotide primer has the sequence 5′ cgc CCC GGG GCCTTCAGCGGTTCCCAG 3′ (SEQ ID NO:50) containing the underlined SmaI restriction site and nucleotides 19 to 36 of the galectin 9 protein coding sequence in FIG. 2A-2B (SEQ ID NO:3).

The 3′ galectin 9 primer has the sequence 5′ cgc GGT ACC CAGGGTTGG AAAGGCTG 3′ (SEQ ID NO:51) containing the underlined Asp718 restriction site followed by nucleotides complementary to position 1029 to 1045 of the galectin 9 protein coding sequence in FIG. 2A-2B (SEQ ID NO:3).

The 5′ galectin 10 oligonucleotide primer has the sequence 5′ cgc CCC GGG TTGTCCTTAAACAACCTAC 3′ (SEQ ID NO:52) containing the underlined SmaI restriction site and nucleotides 124-142 of the galectin 10 protein coding sequence in FIG. 3A-3B (SEQ ID NO:5).

The 3′ galectin 10 primer has the sequence 5′ cgc GGT ACC CACA GAAGCCATTCTG 3′ (SEQ ID NO:53) containing the underlined Asp718 restriction site followed by nucleotides complementary to position 1105-1120 of the galectin 10 protein coding sequence in FIG. 3A-3B (SEQ ID NO:5).

The 3′ galectin 10SV primer has the sequence 5′ CGCGGTACCCTA TGCAACTTTATAAAATATTCC 3′ (SEQ ID NO:54) containing the underlined Asp718 restriction site followed by nucleotides complementary to the 3' end of the galectin 10SV protein coding sequence in FIG. 4A-4B (SEQ ID NO:7).

An efficient signal for initiation of translation in eukaryotic cells, as described by Kozak, M., *J. Mol. Biol. 196*:947-950 (1987) is appropriately located in the vector portion of the construct.

The amplified fragment is isolated from a 1% agarose gel using a commercially available kit ("Geneclean," BIO 101 Inc., La Jolla, Ca.). The fragment then is digested with XbaI and again is purified on a 1% agarose gel. This fragment is designated herein F2.

The vector pA2-GP is used to express the galectin 8, 9, 10, or 10SV protein in the baculovirus expression system, using standard methods, as described in Summers et al, A MANUAL OF METHODS FOR BACULOVIRUS VECTORS AND INSECT CELL CULTURE PROCEDURES, Texas Agricultural Experimental Station Bulletin No. 1555 (1987). This expression vector contains the strong polyhedrin promoter of the *Autographa californica* nuclear polyhedrosis virus (AcMNPV) followed by convenient restriction sites. The signal peptide of AcMNPV gp67, including the N-terminal methionine, is located just upstream of a BamHI site. The polyadenylation site of the simian virus 40 ("SV40") is used for efficient polyadenylation. For an easy selection of recombinant virus the beta-galactosidase gene from *E. coli* is inserted in the same orientation as the polyhedrin promoter and is followed by the polyadenylation signal of the polyhedrin gene. The polyhedrin sequences are flanked at both sides by viral sequences for cell-mediated homologous recombination with wild-type viral DNA to generate viable virus that express the cloned polynucleotide.

Many other baculovirus vectors could be used in place of pA2-GP, such as pAc373, pVL941 and pAcIM1 provided, as those of skill readily will appreciate, that construction provides appropriately located signals for transcription, translation, trafficking and the like, such as an in-frame AUG and

a signal peptide, as required. Such vectors are described in Luckow *et al.*, *Virology 170*:31-39, among others.

The plasmid is digested with the restriction enzyme SmaI and Asp718 and then is dephosphorylated using calf intestinal phosphatase, using routine procedures known in the art. The DNA is then isolated from a 1% agarose gel using a commercially available kit ("Geneclean" BIO 101 Inc., La Jolla, Ca.). This vector DNA is designated herein "V2".

Fragment F2 and the dephosphorylated plasmid V2 are ligated together with T4 DNA ligase. *E. coli* HB101 cells are transformed with ligation mix and spread on culture plates. Bacteria are identified that contain the plasmid with the human galectin 8, 9, 10, or 10SV gene by digesting DNA from individual colonies using XbaI and then analyzing the digestion product by gel electrophoresis. The sequence of the cloned fragment is confirmed by DNA sequencing. This plasmid is designated herein pBacgalectin 8, 9, 10, or 10SV.

5 μg of the plasmid pBacgalectin 8, 9, 10, or 10SV is co-transfected with 1.0 μg of a commercially available linearized baculovirus DNA ("BaculoGold™ baculovirus DNA", Pharmingen, San Diego, CA.), using the lipofection method described by Felgner *et al.*, Proc. Natl. Acad. Sci. USA 84: 7413-7417 (1987). 1μg of BaculoGold™ virus DNA and 5 μg of the plasmid pBacgalectin 8, 9, 10, or 10SV are mixed in a sterile well of a microtiter plate containing 50 μl of serum-free Grace's medium (Life Technologies Inc., Gaithersburg, MD). Afterwards 10 μl Lipofectin plus 90 μl Grace's medium are added, mixed and incubated for 15 minutes at room temperature. Then the transfection mixture is added drop-wise to Sf9 insect cells (ATCC CRL 1711) seeded in a 35 mm tissue culture plate with 1 ml Grace's medium without serum. The plate is rocked back and forth to mix the newly added solution. The plate is then incubated for 5 hours at 27°C. After 5 hours the transfection solution is removed from the plate and 1 ml of Grace's insect medium supplemented with 10% fetal calf serum is added. The plate is put back into an incubator and cultivation is continued at 27°C for four days.

After four days the supernatant is collected and a plaque assay is performed, as described by Summers and Smith, cited above. An agarose gel with "Blue Gal" (Life Technologies Inc., Gaithersburg) is used to allow easy identification and isolation of gal-expressing clones, which produce blue-stained plaques. (A detailed description of a "plaque assay" of this type can also be found in the user's guide for insect cell culture and baculovirology distributed by Life Technologies Inc., Gaithersburg, page 9-10).

Four days after serial dilution, the virus is added to the cells. After appropriate incubation, blue stained plaques are picked with the tip of an Eppendorf pipette. The agar containing the recombinant viruses is then resuspended in an Eppendorf tube containing 200 µl of Grace's medium. The agar is removed by a brief centrifugation and the supernatant containing the recombinant baculovirus is used to infect Sf9 cells seeded in 35 mm dishes. Four days later the supernatants of these culture dishes are harvested and then they are stored at 4°C. A clone containing properly inserted hESSB I, II and III is identified by DNA analysis including restriction mapping and sequencing. This is designated herein as V-galectin 8, 9, 10, or 10SV.

Sf9 cells are grown in Grace's medium supplemented with 10% heat-inactivated FBS. The cells are infected with the recombinant baculovirus V-galectin 8, 9, 10, or 10SV at a multiplicity of infection ("MOI") of about 2 (about 1 to about 3). Six hours later the medium is removed and is replaced with SF900 II medium minus methionine and cysteine (available from Life Technologies Inc., Gaithersburg). 42 hours later, 5 µCi of ^{35}S-methionine and 5 µCi ^{35}S-cysteine (available from Amersham) are added. The cells are further incubated for 16 hours and then they are harvested by centrifugation, lysed and the labeled proteins are visualized by SDS-PAGE and autoradiography.

Example 3: Cloning and Expression in Mammalian Cells

Most of the vectors used for the transient expression of the galectin 8, 9, 10, or 10SV protein gene sequence in mammalian cells should carry the SV40 origin of replication. This allows the replication of the vector to high copy numbers in cells (*e.g.* COS cells) which express the T antigen required for the initiation of viral DNA synthesis. Any other mammalian cell line can also be utilized for this purpose.

A typical mammalian expression vector contains the promoter element, which mediates the initiation of transcription of mRNA, the protein coding sequence, and signals required for the termination of transcription and polyadenylation of the transcript. Additional elements include enhancers, Kozak sequences and intervening sequences flanked by donor and acceptor sites for RNA splicing. Highly efficient transcription can be achieved with the early and late promoters from SV40, the long terminal repeats (LTRs) from Retroviruses, *e.g.* RSV, HTLVI, HIVI and the early promoter of the cytomegalovirus (CMV). However, cellular signals can also be used (*e.g.*, human actin promoter). Suitable expression vectors for use in practicing the present invention include, for example, vectors such as pSVL and pMSG (Pharmacia, Uppsala, Sweden), pRSVcat (ATCC 37152), pSV2dhfr (ATCC 37146) and pBC12MI (ATCC 67109). Mammalian host cells that could be used include, human HeLa, 283, H9 and Jurkart cells, mouse NIH3T3 and C127 cells, Cos 1, Cos 7 and CV1, African green monkey cells, quail QC1-3 cells, mouse L cells and Chinese hamster ovary cells.

Alternatively, the gene can be expressed in stable cell lines that contain the gene integrated into a chromosome. The co-transfection with a selectable marker such as dhfr, gpt, neomycin, hygromycin allows the identification and isolation of the transfected cells.

The transfected gene can also be amplified to express large amounts of the encoded protein. The DHFR (dihydrofolate reductase) is a useful marker to

develop cell lines that carry several hundred or even several thousand copies of the gene of interest. Another useful selection marker is the enzyme glutamine synthase (GS) (Murphy *et al., Biochem. J. 227:*277-279 (1991); Bebbington *et al., Bio/Technology 10:*169-175 (1992)). Using these markers, the mammalian cells are grown in selective medium and the cells with the highest resistance are selected. These cell lines contain the amplified gene(s) integrated into a chromosome. Chinese hamster ovary (CHO) cells are often used for the production of proteins.

The expression vectors pC1 and pC4 contain the strong promoter (LTR) of the Rous Sarcoma Virus (Cullen *et al., Molecular and Cellular Biology*, 438-4470 (March, 1985)) plus a fragment of the CMV-enhancer (Boshart *et al., Cell 41*:521-530 (1985)). Multiple cloning sites, *e.g.*, with the restriction enzyme cleavage sites BamHI, XbaI and Asp718, facilitate the cloning of the gene of interest. The vectors contain in addition the 3′ intron, the polyadenylation and termination signal of the rat preproinsulin gene.

Example 3(a): Cloning and Expression in COS Cells

The expression plasmid, pgalectin 8, 9, 10, or 10SV HA, is made by cloning a cDNA encoding galectin 8, 9, 10, or 10SV into the expression vector pcDNAI/Amp (which can be obtained from Invitrogen, Inc.).

The expression vector pcDNAI/amp contains: (1) an *E. coli* origin of replication effective for propagation in *E. coli* and other prokaryotic cells; (2) an ampicillin resistance gene for selection of plasmid-containing prokaryotic cells; (3) an SV40 origin of replication for propagation in eukaryotic cells; (4) a CMV promoter, a polylinker, an SV40 intron, and a polyadenylation signal arranged so that a cDNA conveniently can be placed under expression control of the CMV promoter and operably linked to the SV40 intron and the polyadenylation signal by means of restriction sites in the polylinker.

A DNA fragment encoding the galectin 8, 9, 10, or 10SV protein and an HA tag fused in frame to its 3′ end is cloned into the polylinker region of the vector so that recombinant protein expression is directed by the CMV promoter. The HA tag corresponds to an epitope derived from the influenza hemagglutinin protein described by Wilson *et al., Cell 37*:767-778 (1984). The fusion of the HA tag to the target protein allows easy detection of the recombinant protein with an antibody that recognizes the HA epitope.

The plasmid construction strategy is as follows. The galectin 8, 9, 10, or 10SV cDNA of the deposited clone is amplified using primers that contain convenient restriction sites, much as described above regarding the construction of expression vectors for expression of galectin 8, 9, 10, or 10SV in *E. coli*. To facilitate detection, purification and characterization of the expressed galectin 8, 9, 10, or 10SV, one of the primers contains a hemagglutinin tag ("HA tag") as described above.

Suitable primers include the following, which are used in this example. The 5′ galectin 8 primer has the sequence 5′cgc CCC GGG gcc atc ATG GCCTATGTCCCCG 3′ (SEQ ID NO:55) containing the underlined SmaI restriction enzyme site followed by nucleotide sequence 52-67 of FIG. 1 (SEQ ID NO:1).

The 3′ galectin 8 primer has the sequence 5′ cgc GGT ACC TTAGAT CTGGACATAGGAC 3′ (SEQ ID NO:56) containing the Asp718 restriction followed by nucleotides complementary to nucleotides 1005-1023 of the galectin 8 coding sequence set out in FIG. 1 (SEQ ID NO:1).

The 5′ galectin 9 primer has the sequence 5′ cgc CCC GGG gcc atc ATGGCCTTCAGCGGTTC 3′ (SEQ ID NO:57) containing the underlined SmaI restriction enzyme site followed by the nucleotide sequence of bases 16-32 of FIG. 2A-2B (SEQ ID NO:3).

The 3′ galectin 9 primer has the sequence 5′ cgc GGT ACC CAGGGTT GGAAAGGCTG 3′ (SEQ ID NO:58) containing the Asp718 restriction followed

by nucleotides complementary to nucleotides 1029-1045 of the galectin 9 coding sequence set out in FIG. 2A-2B (SEQ ID NO:3), including the stop codon.

The 5′ galectin 10 and 10SV primer has the sequence 5′ cgc CCC GGG gcc atc ATGATGTTGTCCTTAAAC 3′ (SEQ ID NO:59) containing the underlined SmaI restriction enzyme site followed by nucleotide sequence 118-135 of FIG. 3A-3B (SEQ ID NO:5).

The 3′ galectin 10 primer has the sequence 5′ cgc GGT ACC CACAG AAGCCATTCTG 3′ (SEQ ID NO:60) containing the Asp718 restriction followed by nucleotides complementary to nucleotides 1105-1120 set out in FIG. 3A-3B (SEQ ID NO:5).

The 3′ galectin 10SV primer has the sequence 5′ CGCGGTACCCTA TGCAACTTTATAAAATATTCC 3′ (SEQ ID NO:54) containing the Asp718 restriction followed by nucleotides complementary to the 3' end of the galectin 10SV coding sequence set out in FIG. 4A-4B (SEQ ID NO:7).

The PCR amplified DNA fragment and the vector, pcDNAI/Amp, are digested with HindIII and XhoI and then ligated. The ligation mixture is transformed into *E. coli* strain SURE (available from Stratagene Cloning Systems, 11099 North Torrey Pines Road, La Jolla, CA 92037), and the transformed culture is plated on ampicillin media plates which then are incubated to allow growth of ampicillin resistant colonies. Plasmid DNA is isolated from resistant colonies and examined by restriction analysis and gel sizing for the presence of the galectin 8, 9, 10, or 10SV-encoding fragment.

For expression of recombinant galectin 8, 9, 10, or 10SV, COS cells are transfected with an expression vector, as described above, using DEAE-DEXTRAN, as described, for instance, in Sambrook *et al.*, MOLECULAR CLONING: A LABORATORY MANUAL, Cold Spring Laboratory Press, Cold Spring Harbor, New York (1989). Cells are incubated under conditions for expression of galectin 8, 9, 10, or 10SV by the vector.

Expression of the galectin 8, 9, 10, or 10SV HA fusion protein is detected by radiolabelling and immunoprecipitation, using methods described in, for

example Harlow *et al.*, ANTIBODIES: A LABORATORY MANUAL, 2nd Ed.; Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York (1988). To this end, two days after transfection, the cells are labeled by incubation in media containing ^{35}S-cysteine for 8 hours. The cells and the media are collected, and the cells are washed and the lysed with detergent-containing RIPA buffer: 150 mM NaCl, 1% NP-40, 0.1% SDS, 1% NP-40, 0.5% DOC, 50 mM TRIS, pH 7.5, as described by Wilson *et al.* cited above. Proteins are precipitated from the cell lysate and from the culture media using an HA-specific monoclonal antibody. The precipitated proteins then are analyzed by SDS-PAGE gels and autoradiography. An expression product of the expected size is seen in the cell lysate, which is not seen in negative controls.

Example 3(b): Cloning and Expression in CHO Cells

The vector pC1 is used for the expression of galectin 8, 9, 10, or 10SV protein. Plasmid pC1 is a derivative of the plasmid pSV2-dhfr [ATCC Accession No. 37146]. Both plasmids contain the mouse DHFR gene under control of the SV40 early promoter. Chinese hamster ovary- or other cells lacking dihydrofolate activity that are transfected with these plasmids can be selected by growing the cells in a selective medium (alpha minus MEM, Life Technologies) supplemented with the chemotherapeutic agent methotrexate. The amplification of the DHFR genes in cells resistant to methotrexate (MTX) has been well documented (see, *e.g.*, Alt, F.W., Kellems, R.M., Bertino, J.R., and Schimke, R.T., 1978, *J. Biol. Chem. 253*:1357-1370, Hamlin, J.L. and Ma, C. 1990, *Biochem. et Biophys. Acta, 1097*:107-143, Page, M.J. and Sydenham, M.A., *Biotechnology 9*:64-68 (1991)). Cells grown in increasing concentrations of MTX develop resistance to the drug by overproducing the target enzyme, DHFR, as a result of amplification of the DHFR gene. If a second gene is linked to the DHFR gene it is usually co-amplified and over-expressed. It is state of the art to develop cell lines carrying more than 1,000 copies of the genes. Subsequently, when the methotrexate is

withdrawn, cell lines contain the amplified gene integrated into the chromosome(s).

Plasmid pC1 contains for the expression of the gene of interest a strong promoter of the long terminal repeat (LTR) of the Rouse Sarcoma Virus (Cullen, *et al.*, Molecular and Cellular Biology, March 1985:438-4470) plus a fragment isolated from the enhancer of the immediate early gene of human cytomegalovirus (CMV) (Boshart *et al.*, *Cell 41*:521-530, 1985). Downstream of the promoter are the following single restriction enzyme cleavage sites that allow the integration of the genes: BamHI, PvuII, and NruI. Behind these cloning sites the plasmid contains translational stop codons in all three reading frames followed by the 3′ intron and the polyadenylation site of the rat preproinsulin gene. Other high efficient promoters can also be used for the expression, *e.g.*, the human β-actin promoter, the SV40 early or late promoters or the long terminal repeats from other retroviruses, *e.g.*, HIV and HTLVI. For the polyadenylation of the mRNA other signals, *e.g.*, from the human growth hormone or globin genes can be used as well.

Stable cell lines carrying a gene of interest integrated into the chromosomes can also be selected upon co-transfection with a selectable marker such as gpt, G418 or hygromycin. It is advantageous to use more than one selectable marker in the beginning, *e.g.*, G418 plus methotrexate.

The plasmid pC1 is digested with the restriction enzyme BamHI and then dephosphorylated using calf intestinal phosphates by procedures known in the art. The vector is then isolated from a 1% agarose gel.

The DNA sequence encoding galectin 8, 9, or 10SV, ATCC Deposit Nos. 97732, 97733 and 97734, respectively, is amplified using PCR oligonucleotide primers corresponding to the 5′ and 3′ sequences of the gene. The galectin 10 sequence is similarly amplified from a human endometrial tumor or human fetal heart cDNA library.

The 5′ galectin 8 primer has the sequence 5′ cgcCCCGGGgccatcATG GCCTATGTCCCCG 3′ (SEQ ID NO:55) containing the underlined SmaI

restriction enzyme site followed by nucleotide sequence 52-67 of FIG. 1 (SEQ ID NO:1). Inserted into an expression vector, as described below, the 5′ end of the amplified fragment encoding human galectin 8 provides an efficient signal peptide. An efficient signal for initiation of translation in eukaryotic cells, as described by Kozak, M., *J. Mol. Biol. 196*:947-950 (1987) is appropriately located in the vector portion of the construct.

The 3′ galectin 8 primer has the sequence 5′ cgc GGT ACC TTAGAT CTGGACATAGGAC 3′ (SEQ ID NO:56) containing the Asp718 restriction followed by nucleotides complementary to nucleotides 1005-1023 of the galectin 8 coding sequence set out in FIG. 1 (SEQ ID NO:1).

The 5′ galectin 9 primer has the sequence 5′ cgc CCC GGG gcc atc ATGGCCTTCAGCGGTTC 3′ (SEQ ID NO:57) containing the underlined SmaI restriction enzyme site followed by the nucleotide sequence of bases 16-32 of FIG. 2A-2B (SEQ ID NO:3). Inserted into an expression vector, as described below, the 5′ end of the amplified fragment encoding human galectin 9 provides an efficient signal peptide. An efficient signal for initiation of translation in eukaryotic cells, as described by Kozak, M., *J. Mol. Biol. 196*:947-950 (1987) is appropriately located in the vector portion of the construct.

The 3′ galectin 9 primer has the sequence 5′ cgc GGT ACC CAGGGTT GGAAAGGCTG 3′ (SEQ ID NO:58) containing the Asp718 restriction followed by nucleotides complementary to nucleotides 1029-1045 of the galectin 9 coding sequence set out in FIG. 2A-2B (SEQ ID NO:3), including the stop codon.

The 5′ galectin 10 and 10SV primer has the sequence 5′ cgc CCC GGG gcc atc ATGATGTTGTCCTTAAAC 3′ (SEQ ID NO:59) containing the underlined SmaI restriction enzyme site followed by nucleotide sequence 118-135 of FIG. 3A-3B (SEQ ID NO:5). Inserted into an expression vector, as described below, the 5′ end of the amplified fragment encoding human galectin 10 provides an efficient signal peptide. An efficient signal for initiation of translation in eukaryotic cells, as described by Kozak, M., J. Mol. Biol. 196:947-950 (1987) is appropriately located in the vector portion of the construct.

The 3′ galectin 10 primer has the sequence 5′ cgcGGTACCCACAG AAGCCATTCTG 3′ (SEQ ID NO:60) containing the Asp718 restriction followed by nucleotides complementary to nucleotides 1105-1120 set out in FIG. 3A-3B (SEQ ID NO:5).

The 3′ galectin 10SV primer has the sequence 5′ CGCGGTACCCTA TGCAACTTTATAAAATATTCC 3′ (SEQ ID NO:54) containing the Asp718 restriction followed by nucleotides complementary to the 3' end of the galectin 10SV coding sequence set out in FIG. 4A-4B (SEQ ID NO:7).

The amplified fragments are isolated from a 1% agarose gel as described above and then digested with the endonucleases SmaI and Asp718 and then purified again on a 1% agarose gel.

The isolated fragment and the dephosphorylated vector are then ligated with T4 DNA ligase. *E. coli* HB101 cells are then transformed and bacteria identified that contained the plasmid pC1 inserted in the correct orientation using the restriction enzyme SmaI. The sequence of the inserted gene is confirmed by DNA sequencing.

Transfection of CHO-DHFR-cells

Chinese hamster ovary cells lacking an active DHFR enzyme are used for transfection. Five μg of the expression plasmid C1 are cotransfected with 0.5 μg of the plasmid pSVneo using the lipofecting method (Felgner *et al.*, *supra*). The plasmid pSV2-neo contains a dominant selectable marker, the gene neo from Tn5 encoding an enzyme that confers resistance to a group of antibiotics including G418. The cells are seeded in alpha minus MEM supplemented with 1 mg/ml G418. After 2 days, the cells are trypsinized and seeded in hybridoma cloning plates (Greiner, Germany) and cultivated from 10-14 days. After this period, single clones are trypsinized and then seeded in 6-well petri dishes using different concentrations of methotrexate (25 nM, 50 nM, 100 nM, 200 nM, 400 nM). Clones growing at the highest concentrations of methotrexate are then transferred

to new 6-well plates containing even higher concentrations of methotrexate (500 nM, 1 μM, 2 μM, 5 μM). The same procedure is repeated until clones grow at a concentration of 100 μM.

The expression of the desired gene product is analyzed by Western blot analysis and SDS-PAGE.

Example 4: Tissue distribution of protein expression

Northern blot analysis is carried out to examine galectin 8, 9, 10, or 10SV gene expression in human tissues, using methods described by, among others, Sambrook *et al.*, cited above. A cDNA probe containing the entire nucleotide sequence of the galectin 8, 9, 10, or 10SV protein (SEQ ID NO:1, 3, 5, or 7, respectively) is labeled with ^{32}P using the *redi*prime™ DNA labeling system (Amersham Life Science), according to manufacturer's instructions. After labeling, the probe is purified using a CHROMA SPIN-100™ column (Clontech Laboratories, Inc.), according to manufacturer's protocol number PT1200-1. The purified labeled probe is then used to examine various human tissues for galectin 8, 9, 10, or 10SV mRNA.

Multiple Tissue Northern (MTN) blots containing various human tissues (H) or human immune system tissues (IM) are obtained from Clontech and are examined with labeled probe using ExpressHyb™ hybridization solution (Clontech) according to manufacturer's protocol number PT1190-1. Following hybridization and washing, the blots are mounted and exposed to film at -70°C overnight, and films developed according to standard procedures.

It will be clear that the invention may be practiced otherwise than as particularly described in the foregoing description and examples.

Numerous modifications and variations of the present invention are possible in light of the above teachings and, therefore, are within the scope of the appended claims.

The entire disclosure of all publications (including patents, patent applications, journal articles, laboratory manuals, books, or other documents) cited herein are hereby incorporated by reference.